**Rose Bengal Chloramphenicol Agar**

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| **Product No.** | **Product Category** | **Specification** |
| HCM121 | Dehydrated Culture Medium | 500g/bottle |

**Intended Use**

For the selective enumeration of moulds and yeasts from foods.

**Principle and Interpretation**

Enzymatic digest of soybean meal provides essential growth nutrients. Dextrose is the fermentable carbohydrate. Monopotassium phosphate act as a buffering agents and magnesium is a trace element important for the growth of yeasts and moulds. Rose bengal dye suppresses the development of bacteria and reduces the spreading of moulds. Chloramphenicol has inhibitory action on gram-negative bacteria.

**Formulation**

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| **Ingredients** | **/liter** |
| Enzymatic digest of soybean meal | 5.0g |
| Dextrose | 10.0g |
| Monopotassium phosphate | 1.0g |
| Magnesium sulfate | 0.5g |
| Agar  | 15.0g |
| Rose bengal  | 0.05g |
| Chloramphenicol | 0.1g |
| pH 7.2±0.2 at 25°C |

**Preparation**

Suspend 31.7g in 1 L of distilled water , stirring heated to boiling until completely dissolved, dispensing flask, 121 ℃ autoclave for 15min. Diluted and treated samples

**Quality Control**

Cultural characteristics observed after incubation at 25 - 30°C for 2-5d ,

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| **Quality control strains** | **Growth** | **Colony color** |
| Aspergillus niger ATCC16404 | PR≥0.5 | white mycelia, black spores |
| Candida albicans ATCC10231 | PR≥0.5 | pink coloured colonies  |
| *Escherichia coli* ATCC25922 | Total inhibition  | - |

**Sorage and Shelf Life**

Keep container tightly closed, store in a cool, dry place, away from bright light. Storage period of 3 years.

**Precautions**

1. When weighing the dehydrated medium, please wear masks to avoid causing respiratory system discomfort

2. Keep container tightly closed after using to prevent clumping.

**Waste Disposal**

Microbiological contamination was disposed by autoclaving at 121°C for 30 minutes.

**Revision**

On June 14, 2024

**References**

1. Smith, N. R. and V. T. Dawson. 1944. The bacteriostatic action of rose bengal in media used for the

plate counts of soil fungi. Soil Sci. 58:467-471.

2. Marshall, R. T. (ed.). 2004. Standard methods for the microbiological examination of dairy products,

17th ed. American Public Health Association, Washington, D.C.

3. Eaton, A. D., L. S. Clesceri, and A. E. Greenberg (eds.). 2017. Standard methods for the

examination of water and wastewater, 23rd ed. American Public Health Association, Washington, D.C.